





Department Of Biomedical And Specialty Surgical Sciences Laboratory of Archeo-Anthropology & Forensic Anthropology

# Microscopic Paleo-Pathology

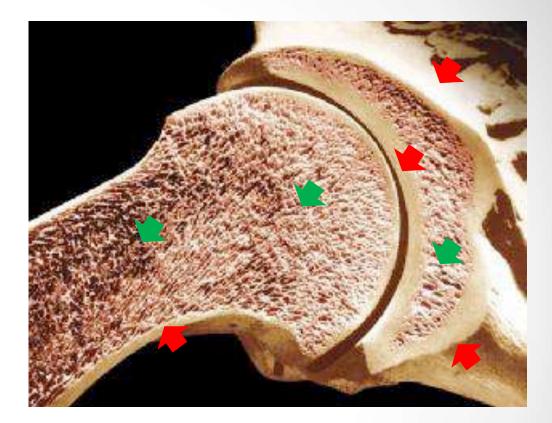
Collection, treatment and preparation of archaeological human remains for histopathological investigation

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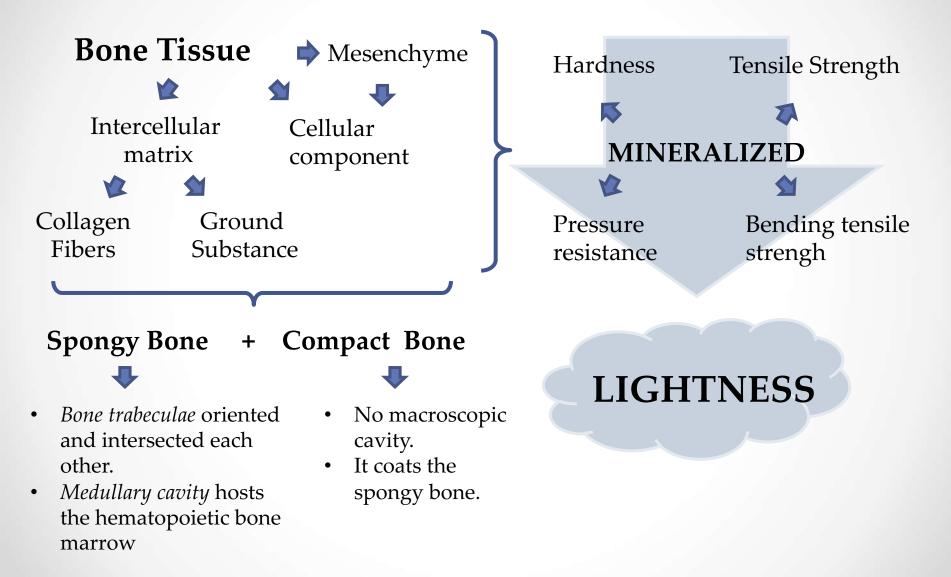
### Introduction

Paleopathology:studiesthe pathological conditionsfound in ancient humanand animal remains.

**Hystology**: studies the microscopic structure of animal and vegetal tissues.



## **Bone Tissue**

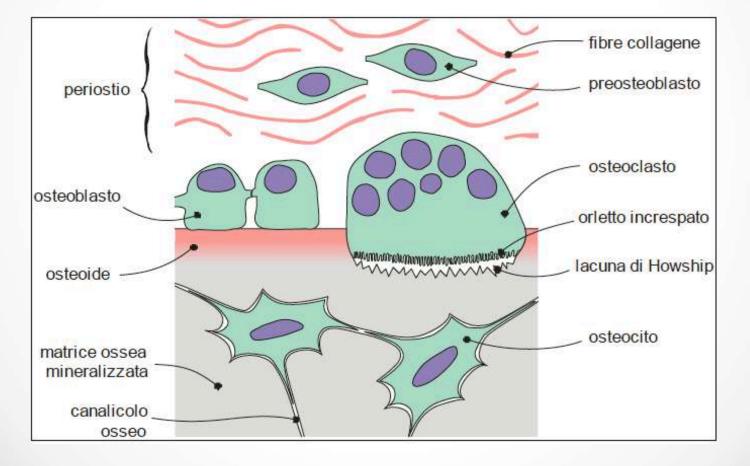


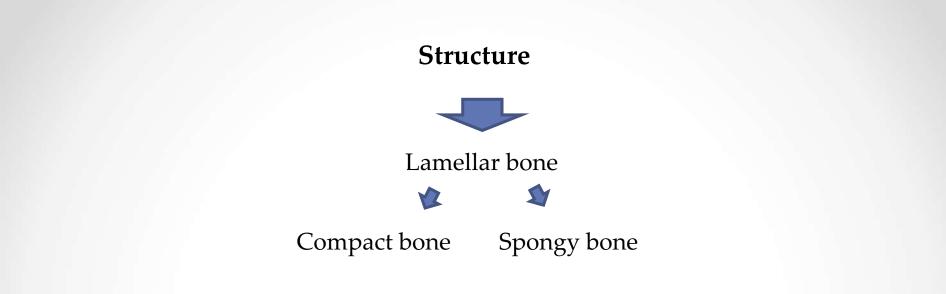
#### **Mineral component**

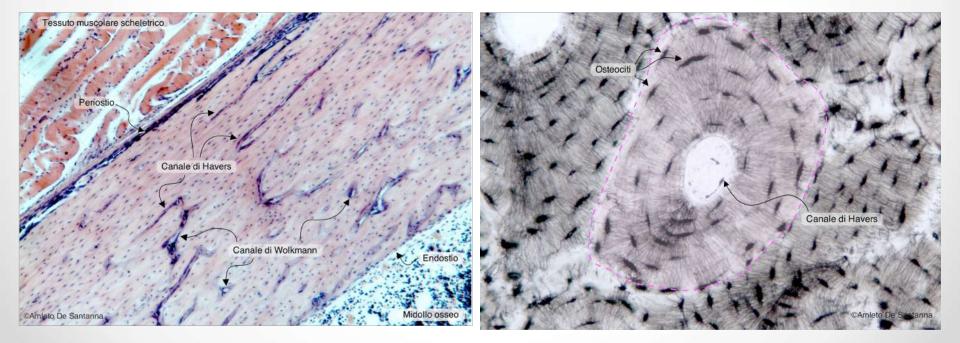
- Calcium phosphate (Hydroxyapatite)
- Calcium carbonate
- Calcium floride
- Magnesium phosphate

### **Cellular components**

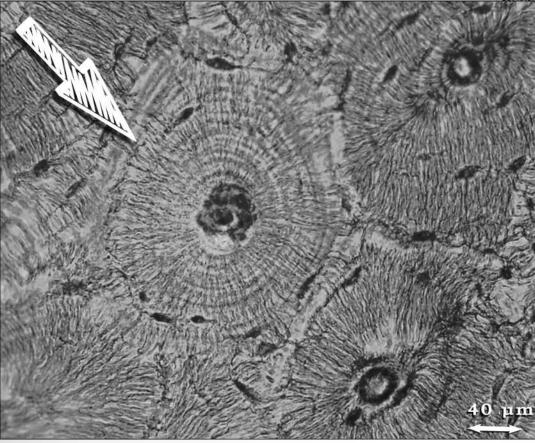
- Pre-Osteoblasts → Osteoblasts
- Osteocytes
- Osteoclasts

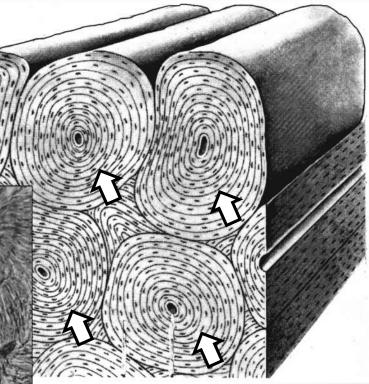




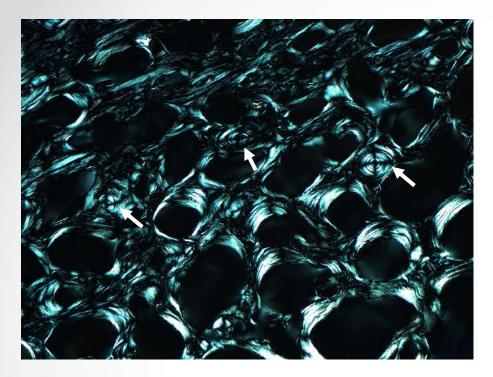


#### **Haversian System**



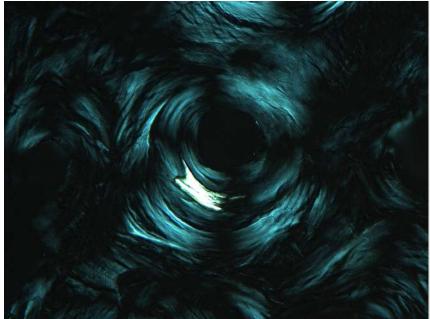


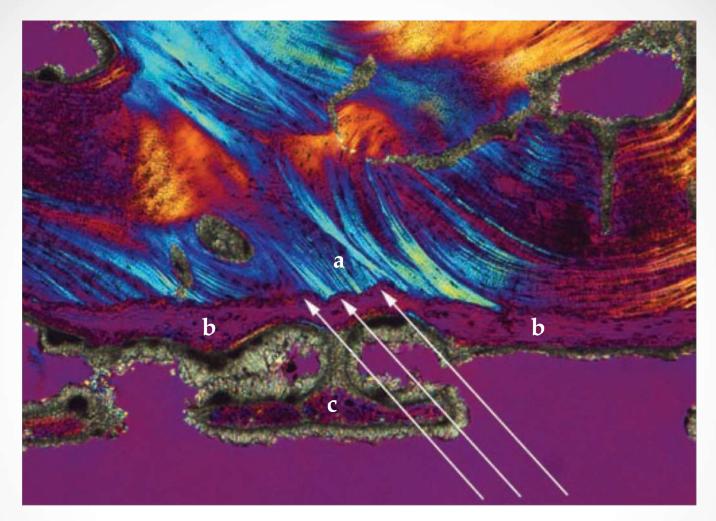
#### Tridimensional drawing of Haversian Systems: Frecce = Haversian System



SSM (Italy), US 81, Middle Adult.
Pariet dx.
Fig. 1) Trabecular bone,
Arrows = Osteons. 25x magnification.

SSM (Italy), US 81, Middle Adult. Pariet dx.. Fig. 2) Osteons. 100x magnification Thin-ground section (50 μm) polarized light microscope.

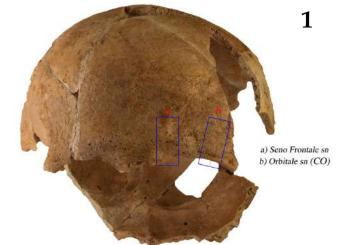




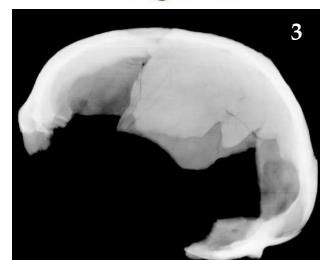
İkiztepe (Turkey), IT-Türk-145, subadult, III millennium BC. Parietal left. CC internal lamina, probable meningeal reaction;  $\mathbf{a} =$  lamellar bone  $\mathbf{b} =$  new bone formation,  $\mathbf{c} =$  conglomerate made by new bone formation, **Arrows** = Howship's lacunae (Thin-ground section (70 µm) Polarized light microscope with polarizing filter and quartz compensator 100 X magnification. © Schultz M.

**Collection, Treatment & Preparation of the archaeological human bones.** 

- 1. Photographic documentation
- 2. Endoscopic/Microscopic documentation
- 3. Radiologic Documentation







Spina VP Ind. 2E

#### **Sampling methods**

Using:

- Manual cutting
- Diamond rotary saw / surgical saw
- Drilling
- Erosion

Measuring and labelling





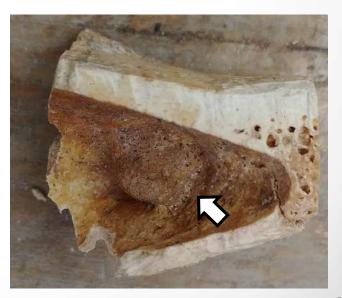


#### Sample's requirement:

- Presence-absence of the lesion
- Possibly cross-cutting
- Minimum sizes: 1 cm<sup>2</sup>
- Maximum sizes: 5 x 7 cm



SSM 81 (Italia) – Sampling steps.



**Collection, Treatment & Preparation of the archaeological human bones.** 

The bones treatment involves some careful steps before the final inclusion in Biodur<sup>©</sup>

- 1. Sediment removal with soft bristle brush
- 2. Removing traces of adhesives with undiluted acetone
- 3. Immersion in Methylene Chloride or in increasing alcoholic solutions







**Collection, Treatment & Preparation of the archaeological human bones.** 

The last step of the treatment consists in the inclusion in resin:

- 1. Immersion of the samples in Epoxy Resin
- 2. Drying of samples in vacuum impregnation system
- 3. Creation of resin discs/cubes with included samples
- 4. Drying of samples in an oven at 25-30°C







**Collection, Treatment & Preparation of the archaeological human bones.** 

The drying phase is followed by two simultaneous phases :



Creation of a polyhedral block with dimensions not exceeding those of the supporting glass.



Preparation of the blocks for longitudinal cutting

Creation of two mirror blocks (50µm and 70µm)



**Regularisation** and Polishing of the surface in contact with the supporting glass.

#### Arrangement on supporting glass

### Thin-ground sections: How to obtain the thin sections.

After the consolidation subsequent to the previous steps , the final procedure shall be followed:

- 1. Reducing block volume
- 2. Thinning to the proper size
- 3. Sanding and Polishing with leather
- 4. Cover with cover glass (max thickness 10µm)
- 5. Labelling of the sample on the slide side









Samplecollectiontable:Differentstagesofcreatingsections.

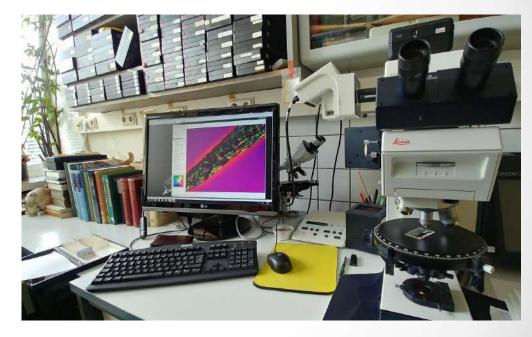
# Microscopy and Paleohistology:

Analysis and possible diagnoses: Some examples

Entire process of creating a batch of Thin-ground sections: 3/4 weeks.

Histological preparations ready for microstructural analysis

Recognition of microscopic structures Comparison with benckmark collection

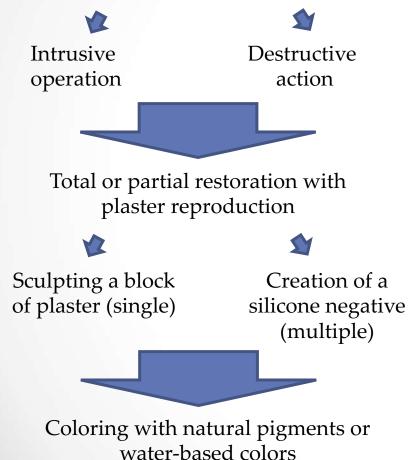


Polarized light microscope Leica DMRXP, Camera Leica DFC 500. Infraview© for digital rendering on PC.

# **Reconstruction work:**

How to repair damage from sampling activities

Tissue collection for histological analysis





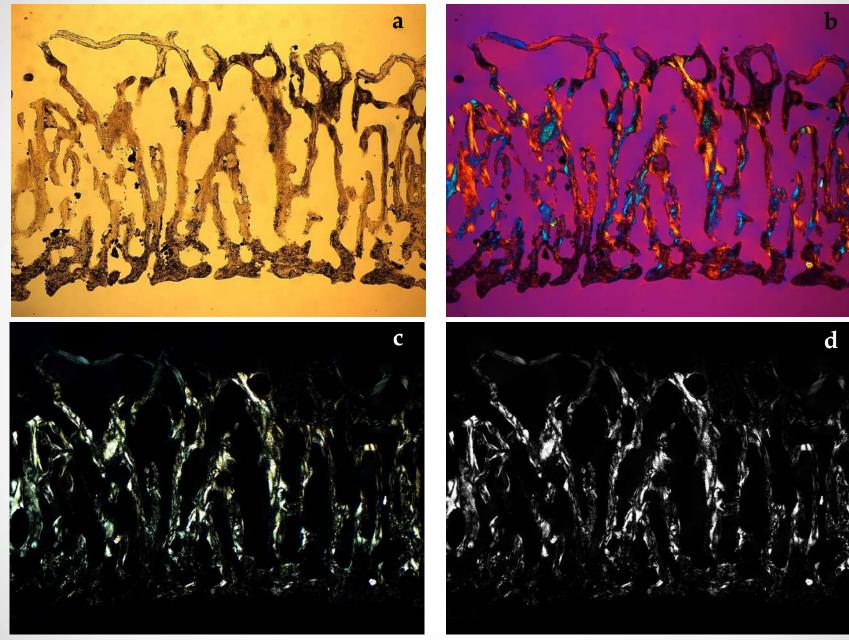
SSM 42 (Italy) – Reconstruction phase.

### Case 1: SSM 4IIa – Subadult Orbital roof





#### SSM 4IIa- LT Orbital Roof



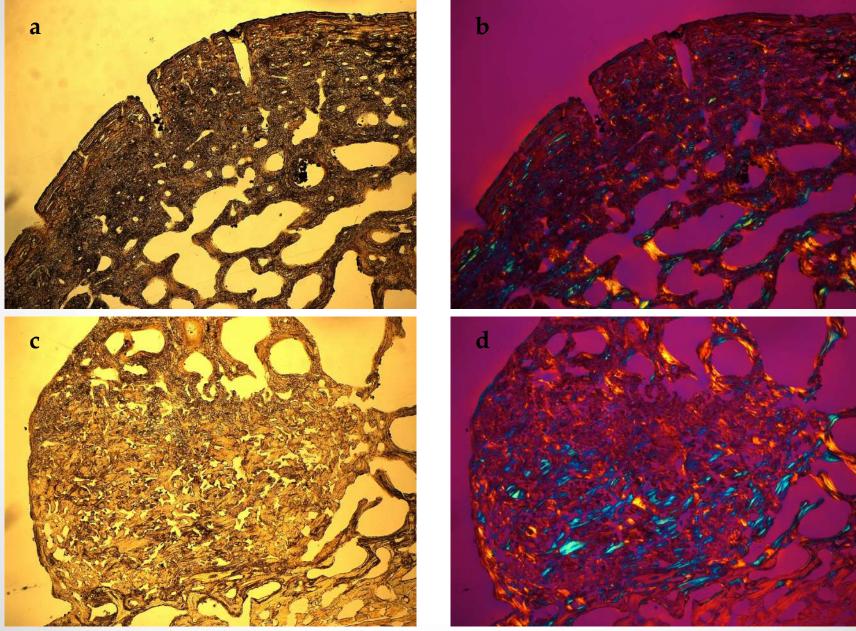
Thin-ground section (50 μm): polarized light microscope using a) transmission light; b) polarizing filter; c)
 polarizing filter & quartz as compensator; d) polarizing filter (de saturated image. 16X magnification

### Case 2: SSM 92 – RT Frontal Sinus + RT Parietal



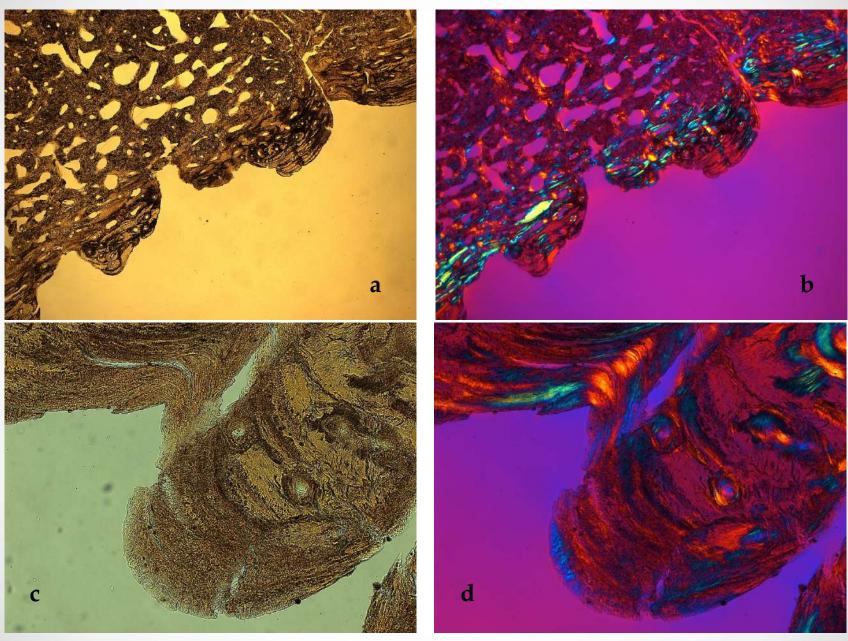


#### SSM 92- RT Frontal Sinus



• Thin-ground section (70 μm): polarized light microscope using **a** - **c**) transmission light; **b** - **d**) polarizing filter and quartz as compensator. 16X magnification

#### SSM 92- RT Parietal



•**Thin-ground section (70 µm):** polarized light microscope using **a**) transmission light; **b**) polarizing filter & quartz as compensator. 16X magnification; **c**) transmission light; **d**) polarizing filter and quartz compensator. Magnification 100X

# Thanks for your attention!

